



DRG[®] Gliadin Deaminated Peptide IgA (Anti) (EIA-5081)



Revised 8 Sept. 2011 rm (Vers. 4.1)



This kit is intended for Research Use Only.

Not for use in diagnostic procedures.

Please use only the valid version of the package insert provided with the kit.

1 INTENDED USE

The Anti-Deamidated Gliadin Peptide (DPG) IgA kit is a solid phase enzyme immunometric assay (ELISA) designed for the measurement of IgA class antibodies directed against deamidated Gliadin peptides (DGP) in human serum or plasma. Anti-Deamidated Gliadin Peptide (DPG) IgA is intended for laboratory use only.

2 PRINCIPLE

Anti-Deamidated Gliadin Peptide (DPG) IgA test is based on the binding of present antibodies in calibrators, controls or prediluted specimen samples on the syntetic deamidated Gliadin peptides (DGP) coated on the inner surface of the wells. After a 30 minutes incubation the microplate is washed with wash buffer for removing non-reactive serum components.

An anti-human-IgA horseradish peroxidase conjugate solution recognizes IgA class antibodies bound to the immobilized antigens. After a 30 minutes incubation any excess enzyme conjugate, which is not specifically bound is washed away with wash buffer.

A chromogenic substrate solution containing TMB is dispensed into the wells. After 15 minutes of incubation the color development is stopped by adding the stop solution. The solutions color changes into yellow. The amount of color is directly proportional to the concentration of IgA antibodies present in the original sample.

3 REAGENTS, MATERIALS AND INSTRUMENTATION

3.1 Reagents and materials supplied in the kit

1. **Anti-DGP Standards S0 – S4;** (5 vials, 1.2 mL each)
Phosphate buffer 0.1 M, NaN₃ < 0.1%, human serum
2. **Control** (2 vials, 1.2 mL each, ready to use)
Phosphate buffer 0.1 M, NaN₃ < 0.1%, human serum
negative and positive control
3. **Sample Diluent** (1 vial, 100 mL)
Phosphate buffer 0.1 M, NaN₃ < 0.1%
4. **Enzyme Conjugate** (1 vial, 15 mL)
Anti h-IgA conjugated with horseradish peroxidase, BSA 0.1%, Proclin < 0.0015%
5. **Coated Microplate** (1 microplate breakable coated with DGP)
6. **TMB-Substrate Solution** (1 vial, 15 mL)
3,3',5,5'-tetramethylbenzidine 0.26 g/L, hydrogen peroxide 0.05%, Proclin < 0.0015%
7. **10X Conc. Wash Solution** (1 vial, 50 mL)
Phosphate buffer 0.2 M, Proclin < 0.0015%



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8. **Stop Solution** (1 vial, 15 mL)
Sulfuric acid 0.15 M

3.2 Reagents necessary not supplied

Distilled water.

3.3 Auxiliary materials and instrumentation

Automatic dispenser.

Microplates reader (450 nm)

4 WARNINGS

1. This kit is intended for research use only.
2. Use appropriate personal protective equipment while working with the reagents provided.
3. All human source material used in the preparation of standards and controls for this product has been tested and found negative for antibody to HIV 1&2, HbsAg, and HCV. No test method however can offer complete assurance that HIV, HBV, HCV or other infectious agents are absent. Therefore, the Standard and the Controls should be handled in the same manner as potentially infectious material.
4. Material of animal origin used in the preparation of the kit has been obtained from animals certified as healthy and the bovine protein has been obtained from countries not infected by BSE, but these materials should be handled as potentially infectious.
5. Some reagents contain small amounts of Sodium Azide (NaN_3) or Proclin 300^R as preservatives. Avoid the contact with skin or mucosa.
6. Sodium Azide may be toxic if ingested or absorbed through the skin or eyes; moreover it may react with lead or copper plumbing to form potentially explosive metal azides. If you use a sink to remove the reagents, allow scroll through large amounts of water to prevent azide build-up.
7. The TMB Substrate contains an irritant, which may be harmful if inhaled, ingested or absorbed through the skin. To prevent injury, avoid inhalation, ingestion or contact with skin and eyes.
8. The Stop Solution consists of a diluted sulphuric acid solution. Sulphuric acid is poisonous and corrosive and can be toxic if ingested. To prevent chemical burns, avoid contact with skin and eyes.
9. Avoid the exposure of reagent TMB/ H_2O_2 to directed sunlight, metals or oxidants.

5 PRECAUTIONS

1. Please adhere strictly to the sequence of pipetting steps provided in this protocol.
2. All reagents should be stored refrigerated at 2 °C – 8 °C in their original container. Any exceptions are clearly indicated.
3. Allow all kit components and specimens to reach room temperature (22 °C - 28 °C) and mix well prior to use.
4. Do not interchange kit components from different lots. The expiry dates printed on the labels of the box and of the vials must be observed. Do not use any kit component beyond their expiry date.



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5. **WARNING: the conjugate reagent is designed to ensure maximum dose sensitivity and may be contaminated by external agents if not used properly;** therefore, it is recommended to use disposable consumables (tips, bottles, trays, etc.). For divided doses, take the exact amount of conjugate needed and do not re-introduce any waste product into the original bottle. In addition, **for doses dispensed with the aid of automatic and semi-automatic devices,** before using the conjugate, it is advisable to clean the fluid handling system, ensuring that the procedures of washing, deproteinization and decontamination are effective in avoiding contamination of the conjugate; **this procedure is highly recommended when the kit is processed using analyzers which are not equipped with disposable tips.**
6. For this purpose, DRG supplies a separate decontamination reagent for cleaning needles.
7. If you use automated equipment is your responsibility to make sure that the kit has been appropriately tested.
8. The incomplete or inaccurate liquid removal from the wells could influence the assay precision and/or increase the background.
9. It is important that the time of reaction in each well is held constant for reproducible results. Pipetting of samples should not extend beyond ten minutes to avoid assay drift. If more than 10 minutes are needed, follow the same order of dispensation. If more than one plate is used, it is recommended to repeat the dose response curve in each plate
10. Addition of the TMB Substrate solution initiates a kinetic reaction, which is terminated by the addition of the Stop Solution. Therefore, the TMB Substrate and the Stop Solution should be added in the same sequence to eliminate any time deviation during the reaction.
11. Observe the guidelines for performing quality control in medical laboratories by assaying controls and/or pooled sera.
12. Maximum precision is required for reconstitution and dispensation of the reagents.
13. Samples microbiologically contaminated should not be used in the assay. Highly lipemic or haemolysed specimens should similarly not be used
14. Plate readers measure vertically. Do not touch the bottom of the wells.

6 STORAGE CONDITION

Store all the kit reagents at 2 °C – 8 °C. Do not freeze. Reagents are stable until the expiration date when stored and handled as directed.

Unused coated microwell strips should be released securely in the foil pouch containing desiccant and stored at 2 °C – 8 °C.

7 PROCEDURE

7.1 Preparation of the Standard (S0 – S4)

Since no international reference preparation for Anti-DGP antibodies is available, the assay system is calibrated in relative arbitrary units. The standards have approximately the following concentration:

	S0	S1	S2	S3	S4
AU/mL	0	15	30	60	240

7.2 Preparation of the Sample

For determination of Anti-DGP human serum or plasma are the preferred sample matrixes.

All serum and plasma samples have to be prediluted with sample diluent 1 : 100.

Therefore 10 µL of sample may be diluted with 990 µL of sample diluent.



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The specimen donor need not to be fasting, and no special preparations are necessary. Collect blood by venipuncture into vacutainers and separate serum (after clot formation) or plasma from the cells by centrifugation.

Samples may be stored refrigerated at 2-8 °C for at least 5 days. For longer storage of up to six months samples should be stored frozen at -20°C. To avoid repeated thawing and freezing the samples should be aliquoted.

Neither Bilirubin nor Hemolysis have significant effect on the procedure.

The Controls are ready to use.

7.3 Preparation of the Wash Solution

Dilute the contents of each vial of the buffered wash solution concentrate (10x) with distilled water to a final volume of 500 ml prior to use.

For smaller volumes respect the 1:10 dilution ratio.

The diluted wash solution is stable for 30 days at 2 °C – 8 °C.

7.4 Procedure

Allow all reagents to stand at room temperature (22 °C - 28 °C).

As it is necessary to perform the determination in duplicate, prepare two wells for each of the five points of the standard curve (S0-S4), two for each control, two for each sample and one for Blank.

Reagent	Standard	Sample or Control	Blank
Standard S0-S4	100 µL		
Controls		100 µL	
Diluted Sample		100 µL	
Incubate 30 minutes at room temperature (22 °C - 28 °C). Remove the contents from each well, wash the wells three times with 300 µL diluted wash solution			
Conjugate	100 µL	100 µL	
Incubate 30 minutes at room temperature (22 °C - 28 °C). Remove the contents from each well, wash the wells three times with 300 µL diluted wash solution.			
TMB substrate	100 µL	100 µL	100 µL
Incubate 15 minutes in the dark at room temperature (22 °C - 28 °C).			
Stop solution	100 µL	100 µL	100 µL
Shake the microplate gently Read the absorbance (E) at 450 nm against Blank.			



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8 RESULTS

8.1 Standard curve

For Anti-DGP IgA a 4-Parameter-Fit with lin-log coordinates for optical density and concentration is the data reduction method of choice. Smoothed-Spline Approximation and log-log coordinates are also suitable.

However, we recommend using a Lin-Log curve.

First calculate the averaged optical densities for each calibrator well. Use lin-log graph paper and plot the averaged optical density of each calibrator versus the concentration. Draw the best fitting curve approximating the path of all calibrator points. The calibrator points may also be connected with straight line segments. The concentration of unknowns may then be estimated from the calibration curve by interpolation.

Typical Results (example only)

The figures below show typical results for For Anti-DGP IgA. These data are intended for illustration only and should not be used to calculate results from another run.

N	OD1	OD2	Mean OD	Conc. 1	Conc. 2	Mean Conc.	CV %
STD0	0,013	0,009	0,011	0,18	0,00	0,09	141,42
STD1	0,205	0,206	0,206	14,93	15,00	14,96	0,36
STD2	0,401	0,405	0,403	29,89	30,20	30,04	0,73
STD3	0,794	0,770	0,782	60,96	59,01	59,99	2,30
STD4	2,558	2,710	2,634	231,2	249,0	240,1	5,26

9 WASTE MANAGEMENT

Reagents must be disposed off in accordance with local regulations.

10 BIBLIOGRAPHY

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11 TROUBLESHOOTING

ERRORS / POSSIBLE CAUSES / SUGGESTIONS

No colorimetric reaction

1. no conjugate pipetted reaction after addition
2. contamination of conjugates and/or of substrate
3. errors in performing the assay procedure (e.g. accidental pipetting of reagents in a wrong sequence or from the wrong vial, etc.)

Too low reaction (too low ODs)

1. incorrect conjugate (e.g. not from original kit)
2. incubation time too short, incubation temperature too low

Too high reaction (too high ODs)

1. incorrect conjugate (e.g. not from original kit)
2. incubation time too long, incubation temperature too high
3. water quality for wash buffer insufficient (low grade of deionization)
4. insufficient washing (conjugates not properly removed)

Unexplainable outliers

1. contamination of pipettes, tips or containers
2. insufficient washing (conjugates not properly removed)

too high within-run CV%

3. reagents and/or strips not pre-warmed to room temperature prior to use
4. plate washer is not washing correctly (suggestion: clean washer head)

too high between-run CV%

5. incubation conditions not constant (time, temperature)
6. controls and samples not dispensed at the same time (with the same intervals) (check pipetting order)
7. person-related variation

Version_2011-08-16~rm